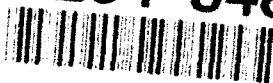


AD-A264 848

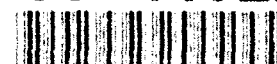


DOCUMENTATION PAGE

Form Approved
OMB No. 0704-0188

1a REPORT SECURITY CLASSIFICATION (U)		1b RESTRICTIVE MARKINGS NA	
2a SECURITY CLASSIFICATION AUTHORITY NA		3 DISTRIBUTION AVAILABILITY OF REPORT Distribution unlimited	
2b DECLASSIFICATION/DOWNGRADING SCHEDULE NA		5 MONITORING ORGANIZATION REPORT NUMBER(S) NA	
4 PERFORMING ORGANIZATION REPORT NUMBER(S) Princeton University		7a NAME OF MONITORING ORGANIZATION Office of Naval Research	
6a NAME OF PERFORMING ORGANIZATION Princeton University	6b OFFICE SYMBOL (If applicable) NA	7b ADDRESS (City, State, and ZIP Code) 800 N. Quincy Street Arlington, VA 22217-5000	
6c ADDRESS (City, State, and ZIP Code) Princeton, NJ 08544		9 PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER N-00014-90-J-1702	
8a NAME OF FUNDING SPONSORING ORGANIZATION Office of Naval Research	8b OFFICE SYMBOL (If applicable) ONR	10 SOURCE OF FUNDING NUMBERS	
8c ADDRESS (City, State, and ZIP Code) 800 N. Quincy Street Arlington, VA 22217-5000		PROGRAM ELEMENT NO 61153N	PROJECT NO RR04108
		TASK NO 441k818	WORK UNIT ACCESSION NO
11 TITLE (Include Security Classification) (U) Lipid Dependent Mechanisms of Protein Pump Activity			
12 PERSONAL AUTHOR(S) Gruner, Sol M.			
13a TYPE OF REPORT Final	13b TIME COVERED FROM 6/90 TO 11/92	14 DATE OF REPORT 29/04/93	
16 SUPPLEMENTARY NOTATION			
17 COSATI CODES		18 SUBJECT TERMS (Continue on reverse if necessary and identify by block number)	
FIELD	GROUP	SUB-GROUP	
08			
		Lipid effect; protein pumps; membrane protein	
19 ABSTRACT (Continue on reverse if necessary and identify by block number)			
<p>The overall objectives of the grant were to investigate the relationship between the activity of membrane proteins, such as protein pumps and channels, and the elastic curvature properties of the imbedding lipid bilayer. The goal was to understand if lipid composition modulates the protein activity via a coupling to the lipid monolayer elastic energies. This involved investigation both of the physical properties of lipid systems and measurement of the effects upon specific proteins. Progress on the following specific objectives are summarized:</p> <ol style="list-style-type: none"> 1) To develop techniques for measuring lipid curvature elastic properties. 2) To investigate if the bilayer spontaneous curvature is regulated in bacterial cells. 3) To correlate ion-pump and channel activity with the spontaneous curvature. 			
20 DISTRIBUTION AVAILABILITY OF ABSTRACT <input checked="" type="checkbox"/> UNCLASSIFIED/UNLIMITED <input type="checkbox"/> SAME AS RPT <input type="checkbox"/> DTIC USERS		21 ABSTRACT SECURITY CLASSIFICATION (U)	
22a NAME OF RESPONSIBLE INDIVIDUAL Dr. Igor Vodyanoy		22b TELEPHONE (Include Area Code) 202-696-4056	22c OFFICE SYMBOL ONR

93-11522



FINAL TECHNICAL REPORT

GRANT #: N00014-90-J-1702

PRINCIPAL INVESTIGATOR: Sol M. Gruner

INSTITUTION: Princeton University

GRANT TITLE: Lipid Dependent Mechanisms of Protein Pump Activity

GRANT PERIOD: 06/01/90 - 11/30/92

Accession	
NTIS GRA&I	
DTIC TAB	
Unannounced	
Justification	
By	
Distribution /	
Availability Codes	
Dist	Avail and/or Special
A-1	

I) Summary

The objectives of the grant were to investigate the relationship between the activity of membrane proteins, such as pumps and channels, and the elastic curvature properties of the imbedding lipid bilayer. The goal was to understand if the lipid composition affects protein activity via a coupling to the bilayer elastic constants. The specific objectives were:

- 1) To develop experimental methods of measuring membrane elastic properties and the interaction with proteins.
- 2) To correlate protein pump activity and channel activity with membrane elastic properties.
- 3) To investigate the relationship between lipid composition and the spontaneous curvature of native membranes.

The first two specific objectives were successfully met, while the third objective is delayed pending receipt of data from collaborators. These results are summarized below.

II) Experimental Methods

The goal here was to use x-ray and neutron diffraction methods to investigate the validity of the physical theory of membrane elastic curvature, as described in detail in Gruner, 1989; 1991; 1992a; 1992b. The primary papers describing the x-ray and neutron diffraction investigations are Turner and Gruner 1992; Turner et al, 1992a; Turner et al, 1992b. The first of these three papers describes the development and application of Fourier methods to reconstruct the structure of the inverse hexagonal phase of certain membrane lipids. The reconstructions are used to show that the density variations in the lipid hydrocarbon diverge as the lamellar-nonlamellar phase transition is approached from high temperature, in agreement with the idea that the lamellar phase represents a state of frustrated curvature. This is important because this curvature is the basis of the coupling of curvature-prone lipid bilayers to proteins imbedded in those bilayers (e.g., see Gruner, 1991).

Turner et al, 1992a describes the combination of x-ray and neutron diffraction to demonstrate that alkanes in the inverted hexagonal phase lattice preferentially occupies hydrocarbon-stressed volumes amongst the lipid chains. This result is significant because it proves that the large effect of short alkanes upon the lamellar-nonlamellar phase transition is due to relaxation of the curvature frustration in curvature-prone bilayer phases, again lending support to the notion that these curvature-frustrated forces are present to exert torques upon imbedded membrane proteins.

The third paper (Turner et al, 1992b) reports on an x-ray diffraction study which is used to extract the elastic constants of lipid layers. The significance of this paper is that it provides numerical estimates of these elastic constants. These, in turn, set limits on the energy available for conformational action on proteins in curvature-frustrated bilayers.

III) Protein Channels and Lipid Curvature

The second specific goal was to use a model protein system to directly demonstrate that lipid spontaneous curvature couples to protein channel function. This was accomplished via the use of alamethicin in a black-lipid membrane (BLM) geometry. These results are described in Keller et al, 1993, which is now in press. The experiment consisted of incorporating alamethicin into BLMs of different lipid compositions and then investigating the electrical channel properties which resulted. The x-ray methods developed earlier were used to characterize the spontaneous curvature of the various lipid compositions. Alamethicin exhibits a series of discrete, well-defined conductance states at fixed transbilayer potential. It was shown that the relative probability of population of the different conductance states varied systematically with the spontaneous curvature. Moreover, when similar, but different lipids were used to make the BLM, compositions with identical curvatures yielded identical effects on the conductance states. These results are highly significant and demonstrate that the physical forces present in curvature-frustrated bilayers coupled to activity in the alamethicin system. Of course, if these forces affect one protein channel, then it is reasonable to assume that other protein channels will also be affected.

IV) Regulation of Native Membrane Lipid Compositions

This goal was to investigate if bacteria regulate the spontaneous curvature of their membranes via control of the lipid composition. This would be reasonable if lipid spontaneous curvature is an important variable affecting membrane proteins vital to the organism. Completion of this goal has been delayed by difficulties in obtaining lipid extracts and some NMR results from our foreign collaborators. Even so, the majority of the work associated with this last objective is completed, as summarized in the letter to Dr. Igor Vodanoy of ONR (dated 9 March 1992). Basically, the results show that the spontaneous curvatures of lipids extracted from mycoplasma membranes cluster tightly, even under growth conditions which result in membrane compositions which are quite varied. Unequivocal

interpretation of the data requires NMR verification of the phase assignments, and calibration of the lipid curvatures against some pure lipid fractions extracted from the lipid mixtures. These steps are being performed, albeit slowly, by the Swedish collaborators who grew the bacteria. We are hopeful of soon obtaining the last of the lipid extracts required to complete interpretation of the data. Even though this information will be obtained beyond the expiration date of the ONR grant, we intend to complete the work, since what remains to be done is primarily analysis and synthesis of the data.

V) Publications, Theses, and Abstracts

1. S.M. Gruner (1989). Stability of lyotropic phases with curved interfaces. Invited feature article. *J. Phys. Chem.* **93**, 7562-7570.
2. O. Narayan, P.T.C. So., D.C. Turner, M.W. Tate, S.M. Gruner and E. Shyamsunder (1990). Volume constriction in a lipid-water liquid crystal using high-pressure x-ray diffraction. *Phys. Rev. A* **42**, 7479.
3. P.T.C. So, M.W. Tate, S.M. Gruner and E. Shyamsunder (1990). X-ray diffraction studies on the effect of alkanols on DOPE-membranes under high pressure. *Biophys. J.* **57**, 274a.
4. S.M. Gruner and E. Shyamsunder (1991). Is the mechanism of general anaesthesia related to lipid membrane spontaneous curvature? *Ann. N.Y. Acad. Sci.* **625**, 685-699.
5. M.W. Tate, E.F. Eikenberry, D.C. Turner, E. Shyamsunder and S.M. Gruner (1991). Nonbilayer phases of membrane lipids. *Chem. Phys. Lipids* **57**, 147-164.
6. S.M. Gruner (1991). Lipid membrane curvature elasticity and protein function. In *Biologically Inspired Physics*, L. Peliti, Ed. (Plenum Press, NY) pp. 127-135.
7. D.C. Turner and S.M. Gruner (1992). X-ray diffraction reconstruction of the inverted hexagonal phase in lipid-water systems. *Biochem. J.* **31**, 1340-1355.
8. D.C. Turner, S.M. Gruner and J.S. Huang (1992). Distribution of decane within the unit cell of the inverted hexagonal phase of lipid-water-decane systems using neutron diffraction. *Biochem. J.* **31**, 1356-1363.
9. S.M. Gruner (1992). Nonlamellar lipid phases. In *The Structure of Biological Membranes*, P.L. Yeagle, ed., (CRC Press, Boca Raton, FL) pp. 211-250.
10. M.W. Tate, E. Shyamsunder, S.M. Gruner and K.L. D'Amico (1992). Kinetics of the lamellar-inverse hexagonal phase transition using time-resolved x-ray diffraction. *Biochem. J.* **31**, 1081-1092.
11. P.T.C. So, S.M. Gruner and E. Shyamsunder (1992). Automated pressure and temperature control apparatus for x-ray powder diffraction studies. *Rev. Sci. Instr.* **63**, 1763-1770.
12. K. Gawrisch, V.A. Parsegian, D.A. Hajduk, M.W. Tate, S.M. Gruner, N.L. Fuller and R.P. Rand (1992). Energetics of the hexagonal-lamellar-hexagonal phase transition sequence in dioleoylphosphatidylethanolamine membranes. *Biochem. J.* **31**, 2856-2864.

13. S.L. Keller, S.M. Bezrukov, S.M. Gruner, I. Vodyanoy and V.A. Parsegian (1992). Relative probability of alamethicin conductance states varies with lipid spontaneous curvature. *Biophys. J.* **61**, a115 (Biophysical Soc. abstract).
14. S.M. Gruner (1992). Nonlamellar lipid phases. *Bull. Amer. Phys. Soc.* **37**, 578-579 (Amer. Phys. Soc. abstract).
15. S.M. Gruner (1992). Coupling between bilayer curvature elasticity and membrane protein activity, in *Advances in Chemistry Series No. 235, Membrane Electrochemistry*, M. Blank and I. Vodyanoy, eds. (ACS Books, Washington, DC, in press).
16. D.C. Turner, Z.-G. Wang, S.M. Gruner, D.A. Mannock and R.N. McElhaney (1992). Structural study of the inverted cubic phases of di-dodecyl alkyl- β -D-glucopyranosyl-rac-glycerol. *J. de Physique* **2**, 2039-2063.
17. P.T.C. So, S.M. Gruner and E. Shyamsunder (1992). High-pressure dilatometer. *Rev. Sci. Instr.* **63**, 5426-5431.
18. S.L. Keller, S.M. Bezrukov, S.M. Gruner, M.W. Tate, I. Vodyanoy and V.A. Parsegian (1993). Probability of alamethicin conductance states varies with nonlamellar tendency of bilayer phospholipids. *Biophys. J.* (in press) (attached).
19. P.T.C. So, S.M. Gruner and E. Shyamsunder (1993). Pressure induced topological transitions in membranes. (Submitted to *Phys. Rev. Lett.*) (attached).
20. A.D. Polcyn, J.T. Gleeson and S.M. Gruner (1993). Microscopic structure of a phospholipid suspension subject to negative pressure. *Bull. Amer. Phys. Soc.* **38**: 405.
21. J.T. Gleeson, E. Shyamsunder and S.M. Gruner (1993). Ice in lamellar lipid phases: A new method for measuring decay of hydration repulsion. *Bull. Amer. Phys. Soc.* **38**: 599.
22. E. Shyamsunder, M. Kriechbaum, F. Osterberg, A.D. Polcyn, V. Skita, P.T.C. So, M.W. Tate and S.M. Gruner (1993). Time-resolved x-ray diffraction of pressure jump induced topological transitions in membranes. *Bull. Amer. Phys. Soc.* **38**: 599.
23. F. Osterberg, S.M. Gruner, M. Kriechbaum, A.D. Polcyn, E. Shyamsunder, V. Skita, P.T.C. So and M.W. Tate (1993). Time-resolved x-ray diffraction of pressure-jump induced relaxations in membranes. *Bull. Amer. Phys. Soc.* **38**: 599.
24. J.T. Gleeson, M.E. Wall, E. Shyamsunder and S.M. Gruner (1993). Freezing and melting of ice in lamellar lipid-water dispersions. *Biophys. J.* **64**: A295.
25. M. Kriechbaum, F. Osterberg, M.W. Tate, E. Shyamsunder, A.D. Polcyn, P.T.C. So, S.M. Gruner and V. Skita (1993). Time-resolved pressure-jump studies on membrane lipids by synchrotron x-ray diffraction. *Biophys. J.* **64**: A296.
26. D.C. Turner (1990). Structural investigations of the inverted hexagonal and inverted cubic phases in lipid-water systems. Ph.D. Thesis, Princeton University, Princeton, NJ.
27. P.T.C. So (1992). High pressure effects on the mesophases of lipid-water systems. Ph.D. Thesis, Princeton University, Princeton, NJ.